

Supplementary Information

The miR528-*D3* module regulates plant height in rice by modulating the gibberellin and abscisic acid metabolisms

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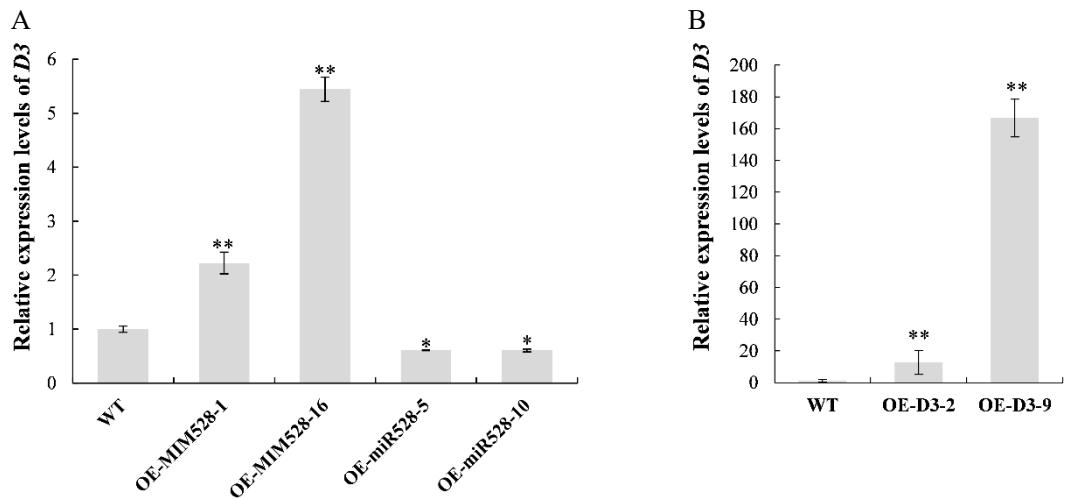


Figure S1. Expression analysis of the *D3* gene in (A) OE-MIM528, OE-miR528 and (B) OE-D3 transgenic plants.

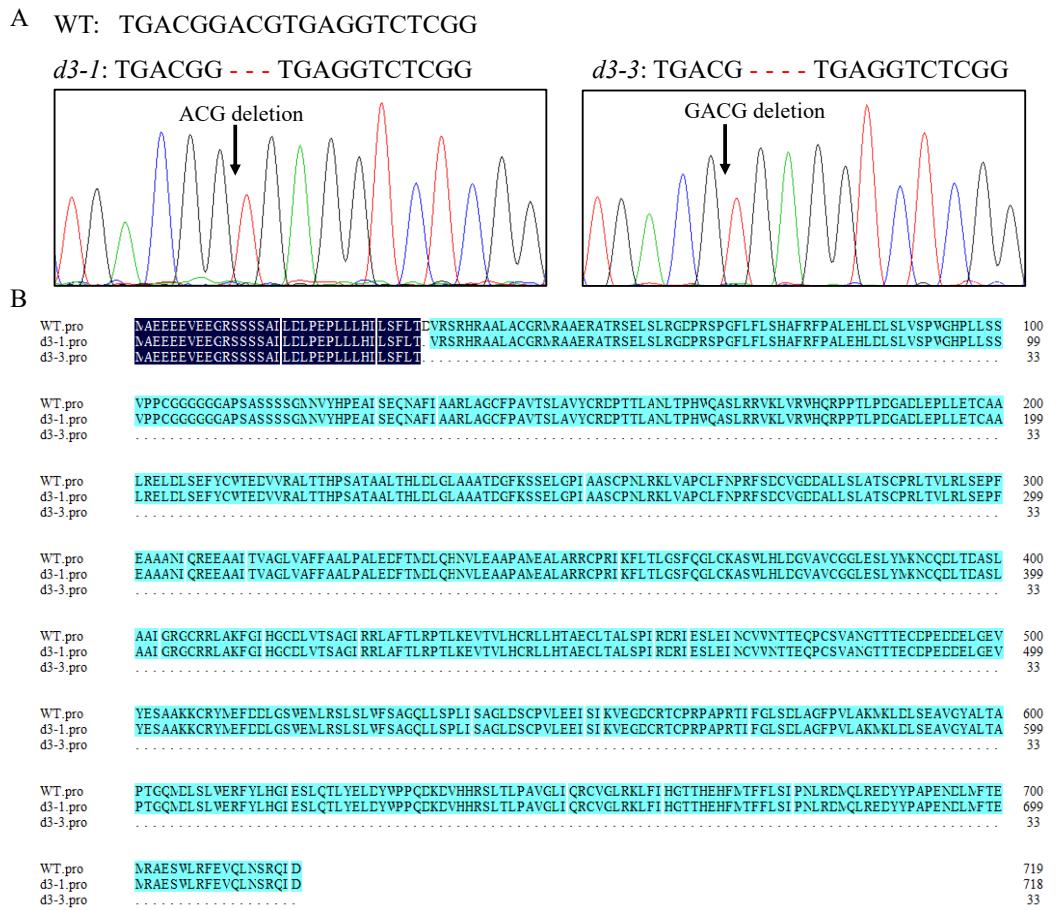


Figure S2. Sequence analysis of *d3* mutants generated by CRISPR/Cas9.

(A) The target sites and sanger sequence of *d3-1* and *d3-3* mutants. (B) Deduced amino acid sequence alignment of *d3* mutants.

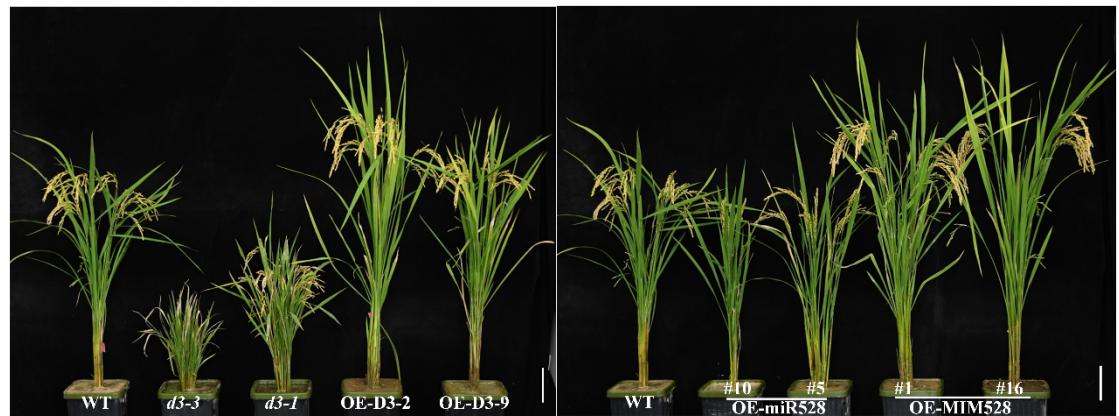


Figure S3. Phenotypes of plant height of different transgenic lines at the maturity stage. Bars, 10 cm.

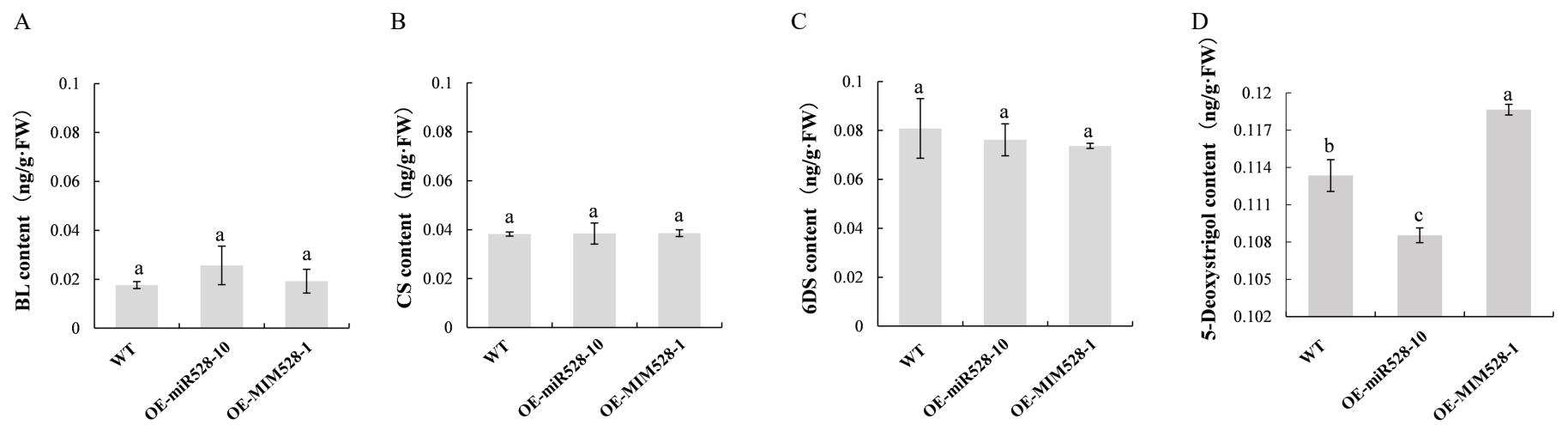


Figure S4. Comparison of the contents of endogenous brassinolide (BL) (A), castasterone (CS) (B), 6-Deoxocastasterone (6-DS) (C), and 5-Deoxystrigol (D) in 31-day-old OE-miR528 and OE-MIM528 transgenic seedlings.

Table S1. Primers used for vector construction and positive identification.

Primer	Sequence
D3cds-F	GCTGGATCCATGGCGGAAGAGGAGGAGGT
D3cds-R	GGACTAGTCATAATCATAACCAACTCACAGGA
U6aF	gccGCCGAGACCTCACGTCCGTC
U6aR	aaacGACGGACGTGAGGTCTCGG
U6bF	gttGGAAGCGGAACGCGTGCAG
U6bR	aaacCTCGCACGCGTTCCGCTTC
SP-F	GCGCGGTGTCATCTATGTTACTA
SP-R	CCCGACATAGATGCAATAACTT
hpt557-F	ACACTACATGGCGTGATTTCAT
hpt557-R	TCCACTATCGGCGAGTACTTCT
d3-F	TTCACCCCCAAATCCCTAAC
d3-R	AGATCGCCTCGGGTGGTAC

Note: D3cds-F/D3cds-R were used for cloning of *D3* gene; U6aF/U6aR, U6bF/U6bR were two specific CRISPR target sites for knockout of *D3* gene; SP-F/SP-R were used for identification of CRISPR/Cas9 tag; hpt557-F/hpt557-R were used for the identification of hygromycin; d3-F/d3-R were used for positive identification of transgenic seedlings.

Table S2. Primers used for qRT-PCR analysis

Gene	Primer	Sequence
<i>OsGA20ox1</i>	OsGA20ox1-FP	TACGCCAGCAGCTTCACGG
	OsGA20ox1-RP	TCCATCAGCTCCAGCGACA
<i>OsGA20ox2</i>	OsGA20ox2-FP	GCAACTACTACCCGCCAT
	OsGA20ox2-RP	CAGGCAGCTCTTATACCTCC
<i>OsGA20ox3</i>	OsGA20ox3-FP	CGCTCACCTCTTCTCAACC
	OsGA20ox3-RP	AGCCATTCTTTGCTTGATCCA
<i>OsGA20ox4</i>	OsGA20ox4-FP	CGCTTCGTGACAAACCTC
	OsGA20ox4-RP	CTGTCCTCGAAGAACTCCC
<i>OsGA3ox1</i>	OsGA3ox1-FP	GATCTTCCATGTGCTCACC
	OsGA3ox1-RP	GAATCATGCTAACGCCGAT
<i>OsGA3ox2</i>	OsGA3ox2-FP	TCTCCAAGCTCATGTGGTCCGAGGGCTA
	OsGA3ox2-RP	TGGAGCACGAAGGTGAAGAAAGCCCGAGT
<i>OsGA2ox1</i>	OsGA2ox1-FP	CGAGAAACGATGTGGAAGGGCTACAGG
	OsGA2ox1-RP	TGGCTCAGGCGGAGTGAGTACATTGTCG
<i>OsGA2ox2</i>	OsGA2ox2-FP	CCCCACATCCCTGACAAGGCTC
	OsGA2ox2-RP	CTATTCATGGTCGTCATCGTCC
<i>OsGA2ox3</i>	OsGA2ox3-FP	TGAGCGCGCTGGTGACGGCGGA
	OsGA2ox3-RP	CTTGATTGTAGGCAGCCTTC
<i>OsGA2ox4</i>	OsGA2ox4-FP	TCGGTGGAGGATAACTTCGGC
	OsGA2ox4-RP	TGGGTTAGCGACAGGTGGTGG
<i>OsGA2ox5</i>	OsGA2ox5-FP	ATGGAGGAGCACGACTACGACT
	OsGA2ox5-RP	TCCTCCATGATCTGCTTCGTGA
<i>OsGA2ox6</i>	OsGA2ox6-FP	GACGACGTGCTTCCTGCGGCTCAA
	OsGA2ox6-RP	CTTCCTGCACCTTCTTCCTGTA
<i>OsGA2ox7</i>	OsGA2ox7-FP	ACGGGAGCTCTACCGAGT
	OsGA2ox7-RP	TCAAATCTGCAGAGCCTGCGTC
<i>OsGA2ox8</i>	OsGA2ox8-FP	GTGCTCGGGGGATGGTGGTGG
	OsGA2ox8-RP	CTTCGTCGCGGCCATCGTTGG
<i>OsGA2ox9</i>	OsGA2ox9-FP	ATGTCGAGGCTGGCCAGGG
	OsGA2ox9-RP	CATACTGAGGAAATTACTGAGGC
<i>OsGA2ox10</i>	OsGA2ox10-FP	ATGAGACAGCTCCGCCGTCTGG
	OsGA2ox10-RP	TTACGTCGTTGTTGATCGTC
<i>OsGA2ox11</i>	OsGA2ox11-FP	CTCCGATCCAACGACACCTCT
	OsGA2ox11-RP	AGCCAGCGCCTCGTCTGAT
<i>OsCPS1</i>	OsCPS1-FP	ACGAATTGAGGAGGCAGCATCTATG
	OsCPS1-RP	GAGCAAGTTCTGCATACCCAACTC
<i>OsCPS2</i>	OsCPS2-FP	CTCTTCCATGTGAGCAAAC
	OsCPS2-RP	CATGCTGGTAGACACAACTC
<i>OsKS2</i>	OsKS2-FP	AGATCGTACCGAGCGCACCTCG
	OsKS2-RP	TGACGGCAGGAACCTCCAAACCCATCTC
<i>OsKS5</i>	OsKS5-FP	CAAACCTCACGCTCCCGCAATGTATCG
	OsKS5-RP	GAATTCCAACCCGTTCCAGTGGCAAGG
<i>OsKO2</i>	OsKO2-FP	ATTCTCCCTACCTCAGCTGGTTCC
	OsKO2-RP	CTCTATGAGTGCCTCCCACACTAGCATC
<i>OsKAO</i>	OsKAO-FP	GAGATCGTCGACGCTCATCATGTACC
	OsKAO-RP	AGATGTTGACGCAAGCGAAGTGTCTCGTC
<i>EUI</i>	EUI-FP	GCCTTGCCGGTGGTGGTGGC
	EUI-RP	GAGGGGTCTCTCCCCGTC